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Exploration of the "Traceless" Reductive Ligation of S-Nitrosothiols

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ABSTRACT

$$\begin{array}{c} Ph \\ Ph - P = 0 \\ Me \end{array}$$

The first "traceless" reductive ligation of S-nitrosothiols using phosphine ester/thioester conjugates is reported. Experiments also show that stable thioimidate compounds could be formed in the reaction between S-nitrosothiols and some phosphine—thioester substrates.

S-Nitrosothiol (RSNO) formation on cysteine residues represents an important post-translational modification that transduces nitric oxide (NO)-dependent signals. However, the detection of RSNOs in biological systems is still a challenge due to the lability of the SNO moiety. The analytical deficiencies become evident when it is observed that reported values of the analysis of the same tissue or biological fluid by different research groups cover some orders of magnitude. In our opinion, SNO is a unique functional group that should have distinct reactivity from other biological functional groups. If we can develop new bioorthogonal reactions specifically targeting SNO and converting unstable S-nitrosothiols to stable products, such a reaction would hold considerable promise in applications for RSNO detection.

Recently, our laboratory developed a reductive ligation of RSNOs which converts RSNOs to stable sulfenamide products in one step (Scheme 1).⁴ We believe the reaction

mechanism is similar to the Staudinger ligation of azides.⁵ RSNO 1 reacts with 2 equiv of ligation substrate 2 to generate the corresponding phosphine oxide and aza-ylide 3.^{4,6} Then, an intramolecular reaction between the ester and aza-ylide may occur to provide phosphorane 5. Finally, hydrolysis of 5 in the presence of water should furnish the final product 6. This reductive ligation is quite selective for SNO moieties and could potentially be applied for RSNO detection.⁴

Although the ligation reaction using the phosphine—ester substrate shown in Scheme 1 has been demonstrated, a modification in which an amide bond is formed between the two coupling partners to give a product without the phosphine oxide moiety might be even more attractive. It might provide a new way to prepare useful sulfenamide derivatives. Similar to the well-studied traceless Staudinger ligation pioneered by Raines and Bertozzi, we proposed that compounds like 7 should undergo a traceless reductive ligation with RSNOs (Scheme 2). As such, the nucleophilic nitrogen atom gener-

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Scheme 1. Reductive Ligation of RSNOs

ated from the reaction between RSNO and the ligation substrate 7 should attack the carbonyl group. Upon hydrolysis, it should produce the sulfenamide 10^8 and liberate the phosphine oxide 11. Here we report the first example of the "traceless" reductive ligation.

Scheme 2. Traceless Reductive Ligation of RSNOs

Four suitable phosphines (compounds 12–15, Scheme 3) were used to examine the traceless reductive ligation of RSNOs. These compounds were prepared following the procedures reported previously. In each case, intramolecular transfer of the acetyl group from the phosphine to SNO-bearing compound would indicate a successful ligation reaction.

t-Butyl SNO **16** was used as the model in this study (Table 1). Since water is necessary for the ligation process, we screened a number of water-containing solvent systems such as THF/water, dioxane/water, CH₃CN/water, DMF/water, etc. The best conditions were found to be a mixture of THF, CH₃CN, and water (1.5/1.5/1). When *tert*-butyl SNO was treated with **12**, no detectable ligation product was observed. We only isolated corresponding phosphine oxide. The flexibility of the methylene bridge in **12** may reduce the rate of cyclization such that the hydrolysis of aza-ylide intermediate predominated. In contrast, the treatment of *t*-BuSNO with ester phosphines **13a**—**c** led to the desired traceless ligation

Scheme 3. Traceless Reductive Ligation Substrates

products 17a-c in moderate yield (35-65%). Thioester 14 is the most efficient substrate for the traceless Staudinger ligation discovered by Raines et al. However, the reaction between t-BuSNO and 14 only provided the ligation product in 22% yield. When substrates 13 and 14 were treated with other relatively stable tertiary RSNOs, corresponding ligation products were obtained in similar yields (see the Supporting Information for details). Under the same conditions, no ligation product was observed for unstable primary RSNOs such as BnSNO and S-nitrosocysteine derivatives. We noticed that these traceless ligations were much slower when compared to the original ligation.⁴ It usually takes more than 12 h for the aza-ylide intermediates (or other possible intermediates such as compound 23 in Scheme 4) to be completely consumed (monitored by TLC or ³¹P NMR). Therefore, the hydrolysis of these intermediates may predominate the reaction.

Table 1. Traceless Ligation of t-BuSNO

entry	substrate	product	yield (%)
1	12	-	_
2	13a	S-N Me	58
3	13b	17a O S-N Ph	35
4	13c	17b O S-N Bn 17c O	65
5	14	S-N Me	22
6	15	S-N Me	23
		Ph. P=O NS Me 18	59

Interestingly, when thioester **15** was used in the reaction, we not only obtained the desired ligation product **17a** in 23%

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yield but also isolated a stable thioimidate product 18 as the major product (59% yield, Table 1, entry 6). Similar imidate compounds were also observed in some Staudinger ligation processes. We noticed the presence of compound 18 under the reaction conditions (THF/CH₃CN/H₂O, with or without phosphine 15) did not lead to any detectable 17a even after 3 days. Therefore, we suspected that sulfenamide 17a was not the hydrolysis product of thioimidate 18. We propose the reaction between RSNO 16 and thioester phosphine 15 involves two different pathways (Scheme 4): in pathway A, the aza-ylide intermediate 19 undergoes desired ligation process to form amidophosphonium salt 21. Then, hydrolysis of the P-N bond of the amidophosphonium salt forms sulfenamide 17a and phosphine oxide 22. In path B, the azaylide intermediate 19 undergoes an intramolecular aza-Wittig reaction¹⁰ to produce the stable thioimidate **18**.

Scheme 4. Proposed Reaction Mechanism of RSNOs

We envisioned the absence of water in the reaction medium might lead to better yield of the thioimidate products. To test this hypothesis, as well as to examine the generality of this process, we tested the reactions of a series of tertiary RSNOs with thioester phosphine **15a** and **15b** in pure THF. As shown in Table 2, in all cases, the thioimidate products were obtained in good to excellent yields. Interestingly, when these tertiary RSNOs were treated with ester phosphine substrates (**13a-c**) in anhydrous THF, we did not observe the formation of any *O*-imidate products.

The thioimidate products obtained here appear to be quite stable. Incubation of them in acidic (such as AcOH) or basic

Table 2. Thioimidate Formation Results

RSNO	phosphine	product/yield
>	15a	Ph Ph-P=O NS- NS- NS- NE Ph Ph-P=O NO-
	15b	NS- S NS- Ph 18a 93%
Ph Ph—SNO Ph	15a	Ph Ph Ph NS Ph Ph Me 23a 79%
	15b	Ph Ph-P=O NS-Ph Ph
SNO	15a	23b 75% Ph Ph P = 0 NS Me 24a 87%
	15b	Ph. P=O S NS Ph
AcHN OMe	e 15b	24b 94% NHAc Ph—P=O NS—CO ₂ Me Ph—25 76%

(such as LiOH) H₂O/CH₃OH mixtures at 25 °C for 24 h does not lead to the formation of any detectable sulfenamide products, nor was any decomposition observed. However, it is possible to convert them to sulfenamides under some conditions. For example, the treatment of compound **18** with CuCl₂¹¹ in the presence of water led to sulfenamide **17a** in excellent yield (Scheme 5).

Scheme 5. Hydrolysis of Thioimidates

In conclusion, the first example of traceless reductive ligation of RSNOs has been demonstrated. We also found

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that stable thioimidate products could be obtained from the reaction between thioester ligation substrates and RSNOs. Refinements of these preliminary studies may lead to new methods for the detection of *S*-nitrosothiols.

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Supporting Information Available: Synthetic procedures, spectroscopic data, and experimental procedures. This material is available free of charge via the Internet at http://pubs.acs.org. OL802663Q

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